

Comparison of haematological and haemodynamic alterations associated with lidocaine, bupivacaine, and ropivacaine epidural anaesthesia in dogs

Mudasir Ahmad Shah * ^{1a}, Bilal Ahmad Malla * ^{2b}, Prakash Kinjavdekar ¹, Aquil Mohmad ^{3b}, GS Amarpal ¹, Rohit Kumar ¹

¹ Division of Surgery, ICAR-Indian Veterinary Research Institute, Izatnagar, Bareilly, Uttar Pradesh, India

² Division of Veterinary Public Health, ICAR-Indian Veterinary Research Institute, Izatnagar, UP, India

³ Division of Parasitology, ICAR-Indian Veterinary Research Institute, Izatnagar, UP, India

Present address

^a Department of Animal Husbandry, Govt of Jammu & Kashmir, India

^b Department of Veterinary Technology, Higher Education Department, Jammu & Kashmir, India

Article info

Received: 24 August 2025

Received in revised form: 23 September 2025

Accepted: 29 September 2025

Published online: 07 October 2025

Keywords

Dogs

Epidural anaesthesia

Haematological parameters

Blood pressure

* Corresponding authors:

Mudasir Ahmad Shah

Email: bilalmalla938@gmail.com

Bilal Ahmad Malla

Email: syedmudasirshah907@gmail.com

Reviewed by:

Priya Singh

Department of Veterinary Surgery and Radiology,
College of Veterinary Science and Animal Husbandry,
Rewa, MP, India

Rukmani Dewangan

Department of Veterinary Surgery and
Radiology, College of Veterinary Science and
A.H., Anjora, Durg-491 001, Chhattisgarh, India

Abstract

Epidural anaesthesia is one among the most frequently used central neuraxial block techniques because of its simplicity and safety. This study aimed to assess the changes in haematological and haemodynamic parameters induced in dogs by epidurally administered lidocaine, bupivacaine, and ropivacaine together with dexmedetomidine in atropine-midazolam premedicated dogs subjected to elective ovariohysterectomy. A total of twenty-four adult dogs were allocated randomly (n=6) to 4 different groups, namely: A (dexmedetomidine), B (dexmedetomidine with lidocaine), C (dexmedetomidine with bupivacaine), and D (dexmedetomidine with ropivacaine). After a 10 minute premedication period, Dexmedetomidine @ 7 µg/kg, Lidocaine @ 4.4 mg/kg, Bupivacaine @ 2 mg/kg and Ropivacaine @ 2 mg/kg were dispensed into the epidural space. Different haematological and haemodynamic parameters were recorded during the study period. Haemoglobin levels showed a significant decline in Group B at the 30 and 90 minute with respect to baseline values. Packed cell volume, neutrophil and lymphocyte counts showed nonsignificant changes in all the groups relative to baseline. At 90 minutes, Group D exhibited a highly significant reduction ($p < 0.05$) in total leukocyte count compared to the other groups. Groups A, C and D exhibited significant variations in systolic, diastolic blood pressure, mean arterial pressure and haemoglobin oxygen saturation (S_pO_2) at various time points with respect to the baseline values. It was concluded that combining dexmedetomidine with lignocaine for epidural administration does not lead to haematological or haemodynamic instability.

This is an open access article under the CC Attribution license (<http://creativecommons.org/licenses/by/4.0/>)

1. Introduction

Epidural anaesthesia is recognized as one of the most effective analgesic procedures. Due to its simplicity and safety, it is one of the most common approach utilized for central neuraxial blockade. It offers effective intraoperative pain relief and enhances postoperative recovery (Brodner et al. 1999; Tomulic et al. 2023). Moreover, epidural anaesthesia is being used more frequently in high risk surgical procedures as well (Vaisanen et al. 1998) and in the management of critically ill animals (Jones 2000). Its advantages include more effective suppression of the surgical stress response and improved postoperative pain management. A variety of pharmacological agents commonly used for epidural anaesthesia include local anaesthetics (Freire et al. 2010; Odette and Smith 2013), morphine administered either alone or in combination with local anaesthetics or other sedatives (Carregaro et al. 2014) as well as α_2 agonists (xylazine) and ketamine.

Lidocaine is one of the most frequently used agents for epidural

anaesthesia; however, its use is associated with dose dependent side effects on the cardiovascular and central nervous system including hypotension and neurotoxicity (Beasley 1999). Concerns over the cardiotoxicity of bupivacaine eventually led to the development of ropivacaine, a safer, long- acting amino ethylamide local anaesthetic with reduced toxic potential. Dexmedetomidine, a powerful and highly selective α_2 agonist has gained wide clinical adoption due to its sedative, analgesic, sympatholytic, and cardioprotective effects particularly lowered blood pressure and heart rate (Gertler et al. 2001). The individual drug doses and therefore the undesirable side effects may be reduced when two or more drugs are administered in combination providing a balanced epidural analgesia with lesser haemodynamic and haematological after effects (Sekhar et al. 2020). Therefore, the current study was taken to evaluate the haematological and haemodynamic variations induced by epidurally administered lidocaine, bupivacaine and ropivacaine in combination with dexmedetomidine in atropine-midazolam premedicated dogs undergoing elective ovariohysterectomy procedure.

2. Materials and Methods

Between October 2015 and May 2016, a total of 24 healthy, client owned, adult mixed breed dogs with an average body weight of 18.1 ± 2.4 kg were presented for elective ovariohysterectomy at the Division of Surgery, ICAR-Indian Veterinary Research Institute, Izatnagar, Uttar Pradesh, India. Food and water were restricted for 12 and 6 hours, respectively, prior to surgery. Written consent was acquired from the owners before proceeding with the surgical procedure. The 24 dogs were randomly assigned to four groups, with 6 animals in each group ($n = 6$) viz. A (dexmedetomidine), B (dexmedetomidine and lidocaine), C (dexmedetomidine and bupivacaine) and D (dexmedetomidine and ropivacaine).

After ensuring proper control and recording all baseline parameters, the animals were premeditated with atropine sulphate (0.5 mg/ml; Bhavani Pharmaceuticals Pvt. Ltd. India) inoculated intramuscularly at a dosage of 0.04 mg/kg body weight. Five minutes later, sedation was induced using an intravenous injection of midazolam (1 mg/ml; Neon Laboratories Ltd. India) at a dose of 0.7 mg/kg body weight. After achieving adequate sedation 10 minutes post midazolam administration, the dogs were positioned in sternal recumbency with their hind limbs positioned cranially. Epidural anaesthetic agents were then administered at the lumbosacral space (L7–S1) as per the following group protocols:

- Group A received Dexmedetomidine at 7 µg/kg (Dextomid 100 µg/ml, Neon Laboratories Ltd., India)
- Group B received Dexmedetomidine at 7 µg/kg + Lidocaine at 4.4 mg/kg (LOX 2%, Neon Laboratories Ltd., India)
- Group C received Dexmedetomidine at 7 µg/kg + Bupivacaine at 2 mg/kg (ANAWIN 0.5%, Neon Laboratories Ltd., India)
- Group D received Dexmedetomidine at 7 µg/kg + Ropivacaine at 2 mg/kg (ROPIN 0.75%, Neon Laboratories Ltd., India).

1% propofol (Nirfol 1%, Nirlife Limited, India) was administered in all the 4 groups intravenously as an intraoperative supplemental anaesthetic agent, whenever required during the procedure.

Blood samples (2.5 ml) were collected from each animal from the cephalic vein into heparinized vials at three time intervals: baseline (0 minutes), 30 minutes and 90 minutes following epidural drug administration. The haematological parameters analyzed were haemoglobin (Hb), packed cell volume (PCV), total leukocyte count (TLC), Neutrophil and lymphocyte count (%) as per Schalm et al. (1975). The haemodynamic parameters were recorded at the following time intervals: baseline (0 minutes), and subsequently at 5, 10, 20, 30, 45, 60, 75 and 90 minutes after the epidural administration of the drugs. The parameters recorded by a non-invasive approach, were systolic blood pressure (SBP), diastolic blood pressure (DBP), and mean arterial pressure (MAP). The oxygen saturation of haemoglobin (SpO₂) was recorded using a pulse oximeter.

The data were first tested for normal distribution using the Shapiro-Wilk test. To compare mean values across different time points among the groups, Analysis of Variance (ANOVA) followed by Duncan's Multiple Range Test (DMRT) was applied. Within each group, changes over time were assessed using one-way repeated measures analysis, as described by Snedecor and Cochran (1994). Statistical analysis was

performed using SPSS version 20.0 (IBM, IL, USA). Differences were considered statistically significant at p value < 0.05 and highly significant at p value < 0.01 .

3. Results and Discussion

Haemoglobin levels have exhibited a nonsignificant reduction ($P > 0.05$) from baseline in groups A, C and D across all observation periods. In contrast, a significant reduction ($P < 0.05$) was noticed in group B at both 30 minute and 90 minute intervals. Packed cell volume (PCV) has decreased notably ($P < 0.05$) at 30 minutes in groups A and B, and at 90 minutes time interval in group C. Group D exhibited no significant changes ($P > 0.05$) throughout the study. Additionally, no remarkable differences ($P > 0.05$) in PCV and haemoglobin values were found between the groups (Table 1). The decline in Hb and PCV can be linked to shifting of fluids from the extravascular space to intravascular space as a compensatory response to maintain cardiac output under anaesthesia during any surgical procedure (Wagner and Hitchcliff 1991; Lijima et al. 2013). Fluid therapy through intravenous route during the procedure leads to haemodilution which further might have decreased the Hb and PCV (Skarda and Muir 1994; Quispe-Cornejo et al. 2022). Earlier studies have shown that epidural administration of xylazine can lead to a marked reduction in Hb and PCV, likely due to an increase in plasma volume resulting from vasodilation, which causes vascular pooling or sequestration of the blood cells in the spleen and lungs during anaesthesia (Skarda 1996; Papudesi et al. 2025). Similarly, subarachnoid injection of α_2 agonists like xylazine and medetomidine has been associated with decreased Hb and PCV levels in goats (Kinjavdekar et al. 2000), and dogs (Amarpal et al. 1998). Further, the drop in Hb and PCV following epidural administration of ropivacaine has been documented in both healthy and uraemic goats (Singh et al. 2005).

Total leukocyte count (TLC) showed a significant reduction ($P < 0.05$) at 90 minutes in groups B and D compared to their baseline levels. Notably, group D exhibited a highly significant decrease ($P < 0.01$) at 90 min mark in comparison to the other groups (Table 1). This decline in TLC may be linked to the sequestration of circulating leukocytes in the spleen or other reservoirs, likely due to reduced sympathetic tone induced by epidural administration of local anaesthetics. Previous studies involving systemic α_2 agonists have also reported a drop in TLC in dogs (Amarpal et al. 1998). Furthermore, a slight but statistically nonsignificant reduction in TLC following epidural xylazine administration was observed in dogs (Kelawala et al. 1996).

Neutrophil counts showed no significant changes ($P > 0.05$) at any time point in any group compared to baseline values. Slight increase, though non-significant in neutrophil counts were observed, likely reflecting the anaesthetic and surgical stress - induced glucocorticoid release, stimulating neutrophil mobilization (Ivascu et al. 2024). A similar rise in neutrophil levels was noted in buffalo calves following administration of xylazine and lidocaine (Singh et al. 2005). Conversely, slight decreases seen in groups B, C and D at various intervals may result from a blunted stress response either by dexmedetomidine with local anaesthetics or indirectly by sedation and analgesia. Reduced neutrophil counts have also been reported in dogs treated with α_2 agonists (Amarpal et al. 1998). Lymphocyte counts in groups A and D appeared to inversely correlate with neutrophil levels, declining whenever neutrophils increased (Table 1). Systolic blood pressure (SBP) (Fig. 1a), diastolic blood pressure (DBP) (Fig. 1b) and mean arterial

Table 1. Mean \pm SD values of hematological parameters in different groups at different time intervals

Parameter	Group	Time interval (min)		
		0	30	90
^a Hb (g/dl)	A	110.2 \pm 12.89	101.6 \pm 5.22	98.6 \pm 5.46
	B	119.2 \pm 18.47	113 \pm 16.58*	100.4 \pm 20.07*
	C	114.4 \pm 15.06	101.6 \pm 22.24	103.6 \pm 26.13
	D	120.8 \pm 12.93	118.8 \pm 12.38	113.4 \pm 6.39
^b PCV	A	0.44 \pm 0.05	0.39 \pm 0.05*	0.36 \pm 0.08
	B	0.46 \pm 0.04	0.42 \pm 0.06*	0.40 \pm 0.07
	C	0.44 \pm 0.04	0.38 \pm 0.07	0.38 \pm 0.07*
	D	0.47 \pm 0.03	0.46 \pm 0.02	0.44 \pm 0.04
^c TLC ($\times 10^9/L$)	A	7.5 \pm 1.61	6.8 \pm 2.03	6.41 \pm 1.59 ^B
	B	9.8 \pm 2.18	10.8 \pm 3.95	7.42 \pm 1.68* ^B
	C	8.45 \pm 3.49	7.99 \pm 3.89	6.44 \pm 1.33 ^B
	D	8.26 \pm 1.58	7.28 \pm 1.57	3.91 \pm 1.76** ^A
Neutrophils (%)	A	64.2 \pm 8.53	66 \pm 8.37	64.6 \pm 5.81
	B	62.4 \pm 10.14	57.6 \pm 13.65	66.8 \pm 10.01
	C	62.8 \pm 6.61	63.4 \pm 5.27	61 \pm 6.08
	D	62.4 \pm 10.71	65.4 \pm 6.54	58.2 \pm 9.96
Lymphocytes (%)	A	28 \pm 5.87	26.4 \pm 7.13	30.8 \pm 5.54
	B	30.4 \pm 9.81	26 \pm 11.02	28 \pm 7.48
	C	26.4 \pm 3.36	29 \pm 5.79	29 \pm 5.34
	D	30 \pm 10.68	26.2 \pm 3.9	34.4 \pm 11.61

^a Hemoglobin, ^b Packed cell volume, and ^c Total leucocyte count

Values with Superscripts A and B among groups represent significant difference ($p < 0.05$) at corresponding intervals. *Significantly different from base value ($p < 0.05$) and **Very significantly different from base value ($p < 0.01$)

pressure (MAP) (Fig. 1c) initially rose in all groups, followed by a gradual decline relative to baseline values over the course of the study.

Local anaesthetics produce a non-selective blockade of sensory, sympathetic and motor nerve fibers. The resulting sympathetic block leads to vasodilation in the anaesthetized regions, which can cause dose - dependent hypotension, a frequently observed cardiovascular complication in animals (Torske and Dyson 2000). However, in the current study, the epidural co-administration of dexmedetomidine with local anaesthetics in groups B, C and D, along with the systemic administration of atropine across all groups might have counteracted the hypotensive effects of local anaesthetics. α -2 agonists are known to cause a biphasic cardiovascular response - initially inducing peripheral vasoconstriction with elevated blood pressure, followed by reduced sympathetic outflow and blood pressure. A similar pattern was observed in the current study. Anticholinergic agents like atropine and glycopyrrolate have been incriminated to cause tachycardia and hypertension in dogs (Alibhai et al. 1996). The decline in blood pressure following the initial increase may be attributed to the metabolism and systemic clearance of dexmedetomidine and atropine. Additionally, midazolam may have promoted to drop in blood pressure, as it is known to exert minimal effects on cardiac function but has been associated with significant reductions in arterial pressure in dogs (Butola and Singh 2007). Furthermore, the use of supplemental propofol during surgery in the present study may have further given rise to the reduction in blood pressure. When Propofol is used for induction as well as for maintenance of anaesthesia, it lowers the systemic blood pressure, primarily due to peripheral vasodilation, cardiovascular depression and reduced cardiac output (Coates et al. 1987; Claeys et al. 1988; Sebel and Lowdon 1989).

In terms of systolic blood pressure (SBP), groups A, B and C showed statistically nonsignificant changes ($P > 0.05$) relative to their baseline values, while group D exhibited an initial significant increase ($P < 0.05$). Animals in group C demonstrated a significant decrease ($P < 0.05$) in SBP in comparison to the other groups at various intervals (Fig. 1a). This decline in SBP in group C may be linked to reduced myocardial contractility due to suppression of baroreceptor reflexes and

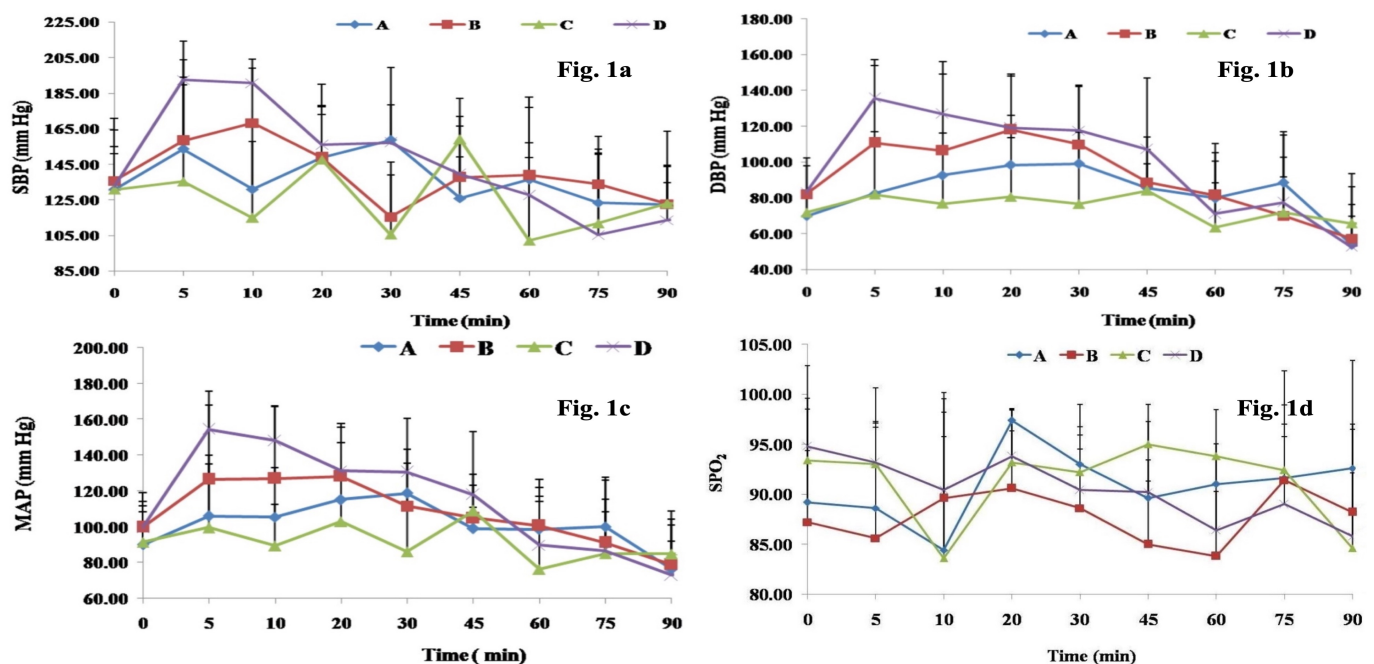


Fig. 1. Mean \pm SD values of (a) Systolic blood pressure (mm Hg), (b) Diastolic blood pressure, (c) Mean arterial pressure (mm Hg), and (d) SpO₂ (%) in different groups at different time intervals

drop in catecholamines due to sympathetic blockade of the adrenal medulla by bupivacaine (Watanabe et al. 1995). Similar findings have been observed where epidural administration of medetomidine in association with ketamine or bupivacaine significantly lowered SBP (Singh et al. 2005). Regarding diastolic blood pressure (DBP) and mean arterial pressure (MAP), both parameters showed a significant increase ($P < 0.05$) initially in groups A and D. However, group A later exhibited a non-significant decline ($P > 0.05$), whereas group D showed a significant decrease ($P < 0.05$) that persisted until the end of the study (Fig. 1b; Fig. 1c). This decline in DBP and MAP is in alignment with the findings of previous studies (Hurley et al. 1991; Duke et al. 2000; Eryilmaz and Günaydin 2011). However, the decrease in group D during the course of study was within clinically acceptable limits.

Mild nonsignificant reduction ($P > 0.05$) in peripheral oxygen saturation (SpO_2) was observed across all groups compared to baseline values. However, group B animals showed a significantly lower SpO_2 ($P < 0.05$) than those in group C at the 45 and 60 minute time intervals (Fig. 1d). The mild non-significant decrease in SpO_2 seen in all groups of dogs may be incriminated to the sedative effects of propofol, which can lead to decreased inspired oxygen levels, hypoventilation, ventilation-perfusion mismatch, diffusion limitations and intrapulmonary shunting (Quandt et al. 1998). Moreover, α_2 agonists have been linked to reduce arterial oxygen partial pressure (PaO_2) in domestic and wild ruminants (Read et al. 2003) and sheep (Kastner 2006) leading to hypoxemia. In a study conducted by Salve et al. (2022) marked reduction in oxygen saturation values under xylazine and tiletamine-zolazepam anaesthesia in dogs has been reported and this can be attributed to lowered respiratory depth due to alpha-adrenergic agonistic action of xylazine. Midazolam has also been linked to lowered SpO_2 levels (Butola and Singh 2007). Similar to the present findings, previous investigations have too reported decreased SpO_2 following the epidural administration of lidocaine in cats, which supports the significant drop observed in animals belonging to group B in comparison to group C at various intervals (DeRossi et al. 2009).

4. Conclusion

Epidural administration of dexmedetomidine with local anaesthetics in atropine- midazolam premedicated dogs does not cause haematological instability and appears safe for elective ovariohysterectomy procedure. However, significant alterations in blood pressure in groups with dexmedetomidine-bupivacaine and dexmedetomidine-ropivacaine are suggestive of the need for better intraoperative monitoring and subsequent measures in cases of any urgency. Among the combinations studied, dexmedetomidine - lignocaine may be considered better than the combination of dexmedetomidine with bupivacaine or ropivacaine as far as haemodynamic stability is concerned.

Declarations

Funding: This study received financial support through the All India Network Program on Diagnostic Imaging and Management of Surgical Conditions in Animals (AINP-DIMSCA) project grants

Conflict of interest: None to declare

Acknowledgements: The authors extend their sincere gratitude to the Director and Joint Director (Research) of ICAR-Indian Veterinary Research Institute, Izatnagar, for their valuable support and for facilitating the necessary infrastructure to carry out this study. They are also thankful to the Indian Council of Agricultural Research (ICAR) for

the financial support and infrastructural assistance provided through the All-India Network Program on Diagnostic Imaging and Management of Surgical Conditions in Animals (AINP-DIMSCA) project.

Ethical approval: As the laboratory animal experiments were not performed in the present research work so, informed consent was acquired from clients of the dogs prior to initiating the elective ovariohysterectomy procedure.

References

- Alibhai HI, Clarke KW, Lee YH, Thompson J. (1996). Cardiopulmonary effects of combinations of medetomidine hydrochloride and atropine sulphate in dogs. *Veterinary Record* 138(1): 11-13. <https://doi.org/10.1136/vr.138.1.11>
- Amarpal, Aithal HP, Kinjavdekar P, Pratap K. (1998). Physiological, haemodynamic and haematological changes due to medetomidine-pethidine induced neuroleptanalgesia in experimental dogs. *The Indian Journal of Animal Sciences* 69(2): 106-108. <https://eprints.icar.org.in/index.php/IJAnS/article/view/20273>
- Beasley VR. (1999). Local Anesthetics. A systems affected approach to veterinary toxicology. Thesis, Department of Veterinary Biosciences, College of Veterinary Medicine, University of Illinois, Urbana, IL, USA. Pp 870-872.
- Brodner B, Mertes B, Van Aken H, Pogatzki E, Buerkle H, Marcus MA, Mollhoff T. (1999). Epidural analgesia with local anesthetics after abdominal surgery: earlier motor recovery with 0.2% ropivacaine than 0.175% bupivacaine. *Anesthesia & Analgesia* 88(1): 128-133. <https://doi.org/10.1213/00005539-199901000-00024>
- Butola V, Singh B. (2007). Midazolam as tranquilizer in dogs. *Indian Veterinary Journal* 84(11): 1141-1145.
- Carregaro AB, Freitas GC, Lopes C, Lukarsewski R, Tamiozzo FS, Santos RR. (2014). Evaluation of analgesic and physiologic effects of epidural morphine administered at a thoracic or lumbar level in dogs undergoing thoracotomy. *Veterinary Anaesthesia and Analgesia* 41(2): 205-211. <https://doi.org/10.1111/vaa.12105>
- Claeys M, Gepts E, Camu F. (1988) Haemodynamic changes during anaesthesia induced and maintained with propofol. *British Journal of Anaesthesia* 60(1): 3-9. <https://doi.org/10.1093/bja/60.1.3>
- Coates DP, Monk CR, Prys-Roberts C, Turtle M. (1987). Hemodynamic effects of infusions of the emulsion formulation of propofol during nitrous oxide anesthesia in humans. *Anesthesia & Analgesia* 66(1): 64-70.
- DeRossi R, Benites AP, Ferreira JZ, Neto JM, Hermeto LC. (2009). Effects of lumbosacral epidural ketamine and lidocaine in xylazine-sedated cats. *Journal of the South African Veterinary Association* 80(2): 79-83. <https://hdl.handle.net/10520/EJC99816>
- Duke T, Caulkett NA, Ball SD, Remedios AM. (2000). Comparative analgesic and cardiopulmonary effects of bupivacaine and ropivacaine in the epidural space of the conscious dog. *Veterinary Anaesthesia and Analgesia* 27(1): 13-21. <https://doi.org/10.1046/j.1467-2995.2000.00001.x>
- Eryilmaz NC, Günaydin B. (2011). A comparison of the effects of intrathecal ropivacaine and bupivacaine during cesarean section. *Turkish Journal of Medical Sciences* 41(2): 219-226. <https://doi.org/10.3906/sag-1008-994>
- Freire CD, Torres ML, Fantoni DT, Cavalcanti RL, Noel-Morgan J. (2010). Bupivacaine 0.25% and methylene blue spread with epidural anesthesia in dog. *Veterinary Anaesthesia and Analgesia* 37(1): 63-69. <https://doi.org/10.1111/j.1467-2995.2009.00493.x>

- Gertler R, Brown HC, Mitchell DH, Silvius EN. (2001). Dexmedetomidine: A novel sedative-analgesic agent. *Baylor University Medical Center Proceedings* 14(1): 13-21. <https://doi.org/10.1080/08998280.2001.11927725>
- Hurley RJ, Arthur RG, Feldman HS, Covino BG, Latka C. (1991). The effects of epinephrine on the anesthetic and hemodynamic properties of ropivacaine and bupivacaine after epidural administration in the dog. *Regional Anesthesia and Pain Medicine* 16(6): 303-308. <https://doi.org/10.1136/rpm-00115550-199116060-00001>
- Ivascu R, Torsin LI, Hostiuc L, Nitipir C, Corneci D, Dutu M. (2024). The surgical stress response and anesthesia: a narrative review. *Journal of Clinical Medicine* 13(10): 3017. <https://doi.org/10.3390/jcm13103017>
- Jones RS. (2001). Epidural analgesia in the dog and cat. *The Veterinary Journal* 161(2): 123-131. <https://doi.org/10.1053/tvj.2000.0528>
- Kästner SBR. (2006). A2-agonists in sheep: a review. *Veterinary Anaesthesia and Analgesia* 33(2): 79-96. <https://doi.org/10.1111/j.1467-2995.2005.00243.x>
- Kelawala NH, Amresh K, Chaudary S, Singh HN, Singh HP. (1996). Effects of epidural xylazine with diazepam premedication in dogs. *Indian Veterinary Journal* 73(5): 552-557.
- Kinjavdekar P, Amarpal GS, Aithal HP, Pawde AM. (2000). Physiologic and biochemical effects of subarachnoidally administered xylazine and medetomidine in goats. *Small Ruminant Research* 38(3): 217-228. [https://doi.org/10.1016/S0921-4488\(00\)00161-9](https://doi.org/10.1016/S0921-4488(00)00161-9)
- Lijima T, Brandstrup B, Rodhe P, Andrijauskas A, Svensen CH. (2013). The maintenance and monitoring of perioperative blood volume. *Perioperative Medicine* 2(1): 9. <https://doi.org/10.1186/2047-0525-2-9>
- Odette O, Smith LJ. (2013). A comparison of epidural analgesia provided by bupivacaine alone, bupivacaine + morphine, or bupivacaine + dexmedetomidine for pelvic orthopedic surgery in dogs. *Veterinary Anaesthesia and Analgesia* 40(5): 527-536. <https://doi.org/10.1111/vaa.12050>
- Papudesi BN, Malayala SV, Regina AC. (2023). Xylazine Toxicity. In: *StatPearls*. StatPearls Publishing, Treasure Island (FL). <https://www.ncbi.nlm.nih.gov/books/NBK594271/>
- Quandt JE, Robinson EP, Rivers WJ, Raffae MR. (1998). Cardiorespiratory and anesthetic effects of propofol and thiopental in dogs. *American Journal of Veterinary Research* 59(9): 1137-1143.
- Quispe-Cornejo AA, Alves da Cunha AL, Njimi H, Mongkolpun W, Valle-Martins AL, Arévalo-López M, Creteur J, Vincent JL. (2022). Effects of rapid fluid infusion on hemoglobin concentration: a systematic review and meta-analysis. *Critical Care* 26(1): 324. <https://doi.org/10.1186/s13054-022-04191-x>
- Read MR. (2003). A review of alpha2 adrenoreceptor agonists and the development of hypoxemia in domestic and wild ruminants. *Journal of Zoo and Wildlife Medicine* 34(2):134-138. [https://doi.org/10.1638/1042-7260\(2003\)034\[0134:AROAAA\]2.0.CO;2](https://doi.org/10.1638/1042-7260(2003)034[0134:AROAAA]2.0.CO;2)
- Salve PD, Thorat MG, Vasantrao R, Raulkar RI, Ingawale MV, Pawar MB, Jadhav AG. (2022). Clinical efficacy of tiletamine-zolazepam and ketamine-diazepam combination on quality of anesthesia for ovariohysterectomy in dog. *Acta Scientific Veterinary Sciences* 4(2): 51-56. <https://doi.org/10.31080/ASVS.2022.04.0308>
- Schalm OW, Jain NC, Carroll EJ. (1975). Normal values in blood morphology with comments on species characteristics in response to disease. In: Schalm OW, Jain NC, Carroll EJ, editors, *Veterinary hematology*. Lea and Febiger, Philadelphia (PA). Pp. 82-218.
- Sebel PS, Lowdon JD. (1989). Propofol: a new intravenous anesthetic. *Anesthesiology* 71(2): 260-277. <https://doi.org/10.1097/0000542-199002000-00035>
- Sekhar KC, Veena P, Kumar RV, Ramayya PJ. (2020). Comparative evaluation of ropivacaine, ropivacaine dexmedetomidine and ropivacaine-clonidine combinations for epidural analgesia in cattle. *Indian Journal of Animal Research* 54(2): 202-208. <https://doi.org/10.18805/Ijar.B-3751>
- Singh K, Kinjavdekar P, Aithal HP, Gopinathan A, Singh GR, Singh T, Pawde AM, Pratap K. (2005). Clinicophysiological and haematobiochemical effects of epidural ropivacaine in uraemic and healthy caprines. *Indian Journal of Veterinary Surgery* 32(1): 11-15.
- Skarda RT. (1996). Local and regional anesthetic and analgesic techniques: dogs. In: Thurmon JC, Tranquilli WJ, Benson GJ, editors, *Lumb and Jones' Veterinary Anesthesia*. Hagerstown, Maryland; Williams & Wilkins, Baltimore, USA. Pp. 434-447.
- Skarda RT, Muir WW. (1994). Caudal analgesia induced by epidural or subarachnoid administration of detomidine hydrochloride solution in mares. *American Journal of Veterinary Research* 55(5): 670-680.
- Snedecor GW, Cochran WG. (1994). Analysis of variance. In: *Snedecor GW, Cochran WG, editors, Statistical methods*. 9th edn. Iowa State University Press, Ames, USA.
- Tomulić Brusich K, Valenčić L, Polonijo Ž. (2023). Physiology and pharmacology of epidurally administered drugs. In: Fyreface-Ogan S, editor, *Epidural administration - New perspectives and uses*. IntechOpen. <http://dx.doi.org/10.5772/intechopen.109116>
- Torske KE, Dyson DH. (2000). Epidural analgesia and anesthesia. *Veterinary Clinics: Small Animal Practice* 30(4): 859-874. [https://doi.org/10.1016/S0195-5616\(08\)70011-1](https://doi.org/10.1016/S0195-5616(08)70011-1)
- Väisänen O, Parviainen I, Ruokonen E, Hippeläinen M, Berg E, Hendolin H, Takala J. (1998). Epidural analgesia with bupivacaine does not improve splanchnic tissue perfusion after aortic reconstruction surgery. *British Journal of Anaesthesia* 81(6): 893-898. <https://doi.org/10.1093/bja/81.6.893>
- Wagner AE, Muir WW, Hinchcliff KW. (1991). Cardiovascular effects of xylazine and detomidine in horses. *American Journal of Veterinary Research* 52(5): 651-657.
- Watanabe Y, Dohi S, Iida H, Ishiyama T, Tashiro T, Shimonaka H. (1995). The effects of systemic bupivacaine on baroreflex sensitivity in dogs. *Masui. The Japanese Journal of Anesthesiology* 44(8): 1097-1101.

Citation

- Shah MA, Malla BA, Kinjavdekar P, Mohmad A, Amarpal GS, Kumar R. (2025). Comparison of haematological and haemodynamic alterations associated with lidocaine, bupivacaine, and ropivacaine epidural anaesthesia in dogs. *Letters in Animal Biology* 05(2): 124 – 128. <https://doi.org/10.62310/liab.v5i2.249>