

Letters in Animal Biology

Journal homepage: www.liabjournal.com

Control of winter ascites in commercial broiler chicken through proprietary formulation

Ibne Ali 1*, Sara Naqvi2

1 Poultry Consultant at consultancy firm "Ali Veterinary Wisdom - Educating the Farmers of the Nation" at New Delhi 2 Life science teacher and advisor at Ahlebiat Trust, New Delhi

Abstract

Article info

Received: 06 November 2022 Received in revised form: 01 December 2022 Accepted: 03 December 2022 Published online: 06 December 2022

Keyword

Ascites Ascitox Metabolic acidosis Reactive oxygen species Mortality Winter

* Corresponding author: Ibne Ali Email: ibnester@gmail.com

Reviewed by:

Meesam Raza

Livestock Production Management, DGCN College of Veterinary and Animal Sciences, CSKHPKV, Palampur - 176062, Himachal Pradesh High growth rates in poultry made them susceptible to various metabolic problems like ascites. The literature has strongly correlated higher metabolic rates with ascites problem in broiler chicken. The workload on the heart and the oxygen demand of the body becomes higher as the growth rate increases. The modern chicken has been genetically selected for higher growth rates, metabolism, and increased feed efficiency which requires more nutrients as well as oxygen. This study assessed the influence of a propriety formulation, Ascitox $-Q_{10}$, on the winter ascites and the consequent livability and growth performance of broiler chicken. The mortality was divided into two stages; M1 (mortality up to 15 days of age) and M2 (mortality after 15 days). The results revealed significantly lower M1 (p<0.05), M2, and overall mortality (p<0.01) in Ascitox supplemented birds compared to control birds. However, the extent of influence of Ascitox was more pronounced on M2 mortality compared to M1 mortality. There was no strain effect on the mortality pattern of birds across different farms under study. In general the farms under study were characterized by spells of sudden deaths overnight. The average weight gain of birds was significantly (p<0.01) higher in Ascitox supplemented birds compared to control birds with no significant effect on their feed intake. However, numerically feed intake of control birds was more than the Ascitox supplemented birds which has led to significantly lower (P<0.01) FCR of Ascitox supplemented birds. In conclusion the supplementation of Ascitox -Q10 is a fruitful nutritional intervention which has controlled the ascites induced mortality in broiler chicken with improved growth performance at commercial farms.

This is an open access article under the CC Attribution-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)

1. Introduction

The growth rate of broiler chicken stocks has drastically risen as a result of consistent selection by poultry breeding companies. As a result, modern chicken consume more feed per unit of time, have better feed efficiency, grow quicker, and have a higher carcass yield than ordinary broiler chicken of 1950s (Havenstein 1994). The contemporary chicken has undergone rigorous selection for faster growth rates, which indirectly leads to faster protein synthesis, the metabolic process requiring more oxygen (Decuypere 2005). Modern strains of broiler chicken are able to achieve market weight in 60% less time than the broiler chicken of 40 years ago. The need for oxygen rises with rapid growth which increases the workload of heart. Thus, fast-growing poultry are more susceptible to ascites development and the degree of ascites and metabolic rate are significantly associated (Al-Zahrani et al. 2019).

However, due to the fact that pulmonary and cardiac capacities of modern broiler chicken strains are guite similar to the old strains, their cardiopulmonary system is forced to operate very close to its physiological limit (Lorenzoni 2006). The lung capacity is not always sufficient to meet the oxygen demands of rapid growth. This results in impaired ability to regulate the energy balance under extreme conditions, such as low ambient temperature or high altitude (Luger et al. 2003). If the lungs of the chicken grow less rapidly than the rest of the body, it could result in hypoxia with consequent ascites (Julian 2000). Pulmonary hypertension syndrome (PHS) or ascites is a metabolic disorder that mostly occurs in fast-growing broiler chicken. High altitude, hypoxia, poor ventilation, low temperature, and fast growth rate are known to be predisposing factors for the incidence of this syndrome (Wideman et al. 1995; Hassanzadeh et al. 1997; Balog 2003).

Pathological findings in ascites induced mortality imply

that the development of a cavity on the exterior surface of the right ventricular wall is the first sign of damage due to pulmonary hypertension. As the injury progresses, it leads to dilation and hypertrophy of the right ventricle resulting in increased blood viscosity, reduced oxygen supply, congestive heart failure, and accumulation of fluids in the abdominal cavity (Biswas 2019; Wideman et al. 2013; Biswas et al. 2012; Ahmed et al. 2013). The two main characteristics of blood that affect its flow in capillaries are plasma viscosity and erythrocyte deformability. And, hypoxia and acidosis significantly impact erythrocyte deformability in a number of clinical situations (Grygorczyk and Orlov 2017). The ability of haemoglobin to bind oxygen is lowered at low blood pH, often known as acidosis. Reduced erythrocyte deformability is a possible contributing factor to pulmonary hypertension and ascites in broiler chickens. The physical characteristics of the blood, primarily haematocrit and plasma viscosity, are important determinants of the rate of oxygen transport in the circulation (Mirsalimi 1991).

According to certain investigations, ascites syndrome may occur as a result of the oxidative stress caused by reactive oxygen species (ROS) (Kalmar et al. 2013). It has been reported that increasing hypoxia caused by insufficient lung function in chickens with ascites is frequently linked to the emergence of PHS (Reeves 1991). The systemic hypoxia may induce cellular hypoxia and increased production of free radicals (Bottje and Wideman 1995). The ROS such as H_2O_2 , superoxide, and hydroxyl ions have potential to induce considerable cell death via lipid peroxidation. They have the ability to oxidize proteins, DNA, and some other tiny molecules inside of a cell, starting a series of negative reactions and harming nearby cells (Halliwell and Gutteridge 1990).

Beyond their role as cellular powerhouses, mitochondria are increasingly being identified as vital players in molecular signal transduction and cell fate determination via reactive oxygen species (ROS). Mitochondria are especially abundant in cardiac tissue; hence, mitochondrial dysregulation and ROS production are thought to contribute significantly to cardiac pathology. Moreover, there is growing appreciation that medical therapies designed to mediate mitochondrial ROS production can be important strategies to ameliorate cardiac disease (Peoples et al. 2019). Recently it has been identified that mitochondrial ability to produce ATP is been diminished by ROS due to the mutations in key genes present in mitochondrial DNA responsible for ATP generation. Depletion of ATP in cardiac myocytes is one of the causes of their inefficiency and eventually failure (Kornfeld et al. 2015).

In the light of above information it can be asserted that ascites and sudden death syndrome are two sides of the same coin. Nevertheless, both the conditions are due to reduced oxygen availability and concomitant cardiac muscle inefficiency which leads to hypertrophy of cardiac muscles and reduced pumping power of heart. This study aimed to assess the role of Ascitox – Q10, a proprietary dietary formulation, in amelioration of ascites and sudden death syndrome in broiler chicken in winter season.

2. Materials and methods

Among different broiler chicken farms located at different locations of same eco-climatic zone in northern India, nine farms showing the cases of ascites were selected for this study. The period of study was from November 2021 to February 2022. Normal winter management practices were followed at all farms and no ascites preventive management was followed at any farm except farm 2, 4, 5, 7, and 8 which were given a dietary proprietary formulation - Ascitox on 2-3-4, 13-14-15, and 23-24-25 day for the amelioration of ascites in birds. The Ascitox was given at the rate of 4 g/L drinking water. Farm 1 to 6 reared and managed Cobb 430Y and farm 7 to 9 reared and managed Skylark Hubbard broiler chicken strains according to the conventional feeding and management practices. Brooding were done in closed house inside brooding guard and under heating source with false ceiling at all the farms (Fig. 1). The mortality from first day to the end of



Fig.1 Brooding condition in farms with brooder guards and false ceiling

rearing period were recorded. Postmortem of dead birds was conducted regularly on all the farms for tentative diagnosis by the available qualified veterinarians. The feed conversion ratio (FCR) and total feed consumption was recorded to access the tentative economic losses due to the ascites in the birds. For convenience of interpretation total mortality was divided into 2 parts; M1 indicating mortality in first 15 days and M2 indicating mortality after 15 days. It was hypothesized that mortality due to ascites will be seen after 15 days of age when metabolism and oxygen requirements are high. Mortality lesions were specially identified during postmortem for the specific pathology associated with ascites in all the birds. Water belly condition in dead birds was given maximum weightage among all the signs associated with ascites (Fig. 2). The four main postmortem findings considered before declaring any mortality due to ascites were water belly with



Fig.2 Specific water belly condition in ascites

plasma like fluid in abdomen, enlarged liver with blood clots over it (Fig. 3), hypertrophy of right ventricle of the heart, and oxidative stress markings on the breast muscles.

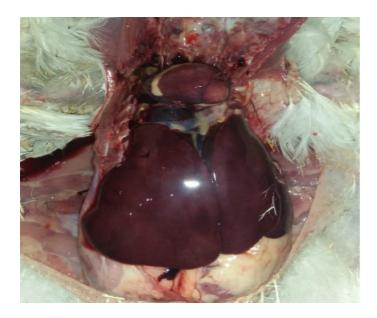


Fig.3 Enlarged blood filled liver and heart; a hallmark of ascites

3. Results and discussion

This study assessed the influence of a propriety formulation, Ascitox , on the winter ascites and the consequent livability and growth performance of broiler chicken. The results in Table 1 revealed significantly lower M1 (p<0.05), M2, and overall mortality (p<0.01) in Ascitox supplemented birds compared to control birds. However, the extent of influence of Ascitox was more pronounced on M2 mortality compared to M1 mortality (Fig. 4). There was no strain effect on the

7.0

mortality pattern of birds across different farms under study.

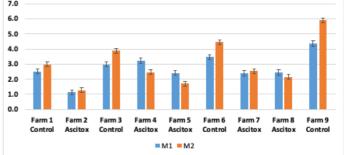


Fig.4 Mortality of birds under the influence of Ascitox (M1: mortality upto 15 days; M2: mortality after 15 days)

Ascites is a common problem of modern day fast growing broiler chicken characterised by fluid accumulation in the abdominal cavity. Because of the disproportionate increase in muscle mass with respect to the normal development of circulatory and respiratory system of modern day broiler chicken the incidence of ascites has increased disproportionately as well (Druyan 2012). It is the condition more commonly witnessed in the winter season because of poor ventilation of the farms due the use of curtains along with the burning of wood, charcoal, and gas to maintain the shed temperature in India. Such activities generate the air contaminants such as ammonia, carbon dioxide, carbon monoxide, dust, and humidity (Aviagen 2009) which creates a hypoxic environment for the birds. This hypoxic environment during the brooding stage of the birds damage their air sacs which makes the respiratory system incompetent to withstand the oxygen demand during the rapid growth phase of birds after 4th week and hence leads to ascites with consequent death. On similar pattern M2 mortality (after 15 days) of birds was higher compared to M1 mortality in this study. Concurrent to this observation it has been reported that hypoxia induced during brooding stage retards the development of lungs, which later on hastens the development of ascites (Wuyi Liu 2016). Furthermore, during the finisher phase of broiler chicken high energy diets are provided to meet the nutritional demands requiring higher oxygen levels for metabolism, and this in turn demands high blood flow through heart and lungs (Dahal 2011). This physiological demand of high blood flow causes hypertension in pulmonary artery which is not handled well by the less flexible blood capillaries of the lungs resulting in effusion of blood fluids in the body cavity.

It has been postulated that broiler chickens with a high metabolic rate may be in a state of metabolic acidosis when fed at full capacity (Julian 1993). Several studies have shown that fast-growing birds have lower blood oxygen concentrations than slow-growing birds; similarly, birds on full feed have lower blood oxygen concentrations than food-deprived birds

Table 1. Mortality pattern in broiler chicken under the influence of Ascitox-Q10									
Parameters	Farm strength	M1	M1 (%)	Farm strength after M1	M2	M2 (%)	Overall mortality (%)		
Control	3068	99	3.22	2969	123	4.15	7.24		
Ascitox	2580	64	2.50	2516	55	2.19	4.61		
SEM	27	3.5	0.17	27	2.9	0.16	0.21		
Significance	-	-	P<0.05	-	-	P<0.01	P<0.01		
Strain Effects									
Cobb 430Y	2778	79	2.84	2700	85	3.13	5.90		
Skylark Hubbard	2833	81	2.87	2752	87	3.16	5.93		
Significance	-	-	P>0.05	-	-	P>0.05	P>0.05		
M1: Mortality upto 15 days; M2: Mortality after 15 days; SEM: Standard error of mean									

(Fedde et al 1998). Therefore, in this study a feed withdrawal of 4 hours was practiced uniformly as a preventive measure in all the farms. In early grower stage of broiler chicken temporary feed restriction potentially reduces incidence of ascites without compromising the growth performance (Acar et al. 1995; Decuypere et al. 2000). On the other hand, limiting the quantity of feed by 40% has also reduced the ascites induced mortality in broiler chicken (Mohammadalipour et al. 2017). In general the farms under study were characterized by spells of sudden deaths overnight (Fig. 5). Earlier studies have also associated ascites with sudden death in well-nourished broiler chicken preceded by brief flapping of their wings for 1-2 minutes (Saki and Hemati 2011). Fast growing young and healthy broiler chicken have been observed to die suddenly while standing, walking, or feeding (Julian 2005).

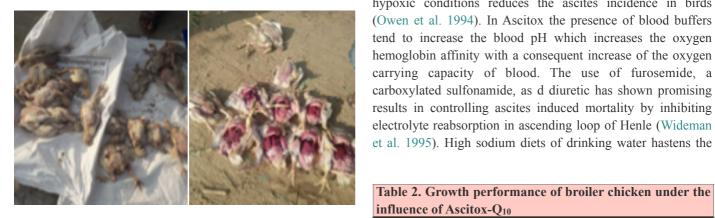


Fig.5 Sudden overnight mortality of birds

This induced hypoxia favors development higher amount of ROS generation in heart muscles and start causing further complications including arrhythmias. Cardiac arrhythmias can occur in broiler chicken as early as 7 days of age, and the incidence rises with age (Olkowski 2007). But nutritional interventions which aimed to correct the metabolic and biochemical disarrangements in the body could reduce the ascites induced mortality in broiler chicken significantly. In this study Ascitox was used as a nutritional intervention whose composition is: CoEnzyme Q₁₀, blood buffers, transmembrane antioxidants, herbal diuretics, herbal antioxidants, sodium ascorbate, feurosemide, and sodium selenite. The results depicting the effects of ascites and ameliorating effects of Ascitox on the growth performance of birds are given in Table 2. The average weight gain of birds was significantly (P < 0.01) higher in Ascitox supplemented birds compared to control birds with no significant effect on their feed intake. However, numerically feed intake of control birds was more than the Ascitox supplemented birds which has led to significantly lower (P<0.01) FCR of Ascitox supplemented birds. Corroborating the results of the present study, modifications in the acid-base balance of the chicken feed under potential hypoxic conditions reduces the ascites incidence in birds (Owen et al. 1994). In Ascitox the presence of blood buffers tend to increase the blood pH which increases the oxygen hemoglobin affinity with a consequent increase of the oxygen carrying capacity of blood. The use of furosemide, a carboxylated sulfonamide, as d diuretic has shown promising results in controlling ascites induced mortality by inhibiting electrolyte reabsorption in ascending loop of Henle (Wideman et al. 1995). High sodium diets of drinking water hastens the

influence of Ascitox-Q ₁₀										
Parameters	Farm strength	Total mortality	Average weight gain (kg)	Average feed intake (kg)	FCR					
Control	3068	222	2.12	3.84	1.81					
Ascitox	2580	119	2.29	3.71	1.62					
SEM	27	3.5	0.07	0.09	0.06					
P value	-	-	P<0.01	P<0.05	P<0.01					
SEM: Standard error of mean										

development of ascites in fast growing broiler chicken (Julian et al. 1990) and use of diuretics reduces sodium and fluid retention which reduces ascites induces mortality.

The supplementation of vitamins, trace minerals, amino acids, and antioxidants like CoEnzyme Q10 have been reported reducing ascites induced mortalities in broiler chicken (Roch 2000; Xiang 2002; Geng et al. 2004) and Ascitox -Q10 used in this study is a balanced mixture of all these nutrients. Reduced pulmonary arterial pressure and reduced ascites index was observed in broiler chicken supplemented with vitamin C and E (Ruiz-Feria 2009). However, supplementing vitamin C and E individually was found to be ineffective in controlling ascites induced mortality of broiler chicken (Khajali and Fahimi 2010; Villar-Patino et al. 2002). This indicates that antioxidants work in a coordinated fashion to provide protection against ROS. The selenium supplementation in the range of 0.15 to 0.30 mg/ kg diet have resulted in significant improvement in ascitesrelated variables in broiler chicken (Ozkan et al. 2007) and nano-selenium at the level of 0.30 mg/kg diet was observed successfully preventing the right ventricular hypertrophy in broiler chicken (Moghaddam et al. 2017). The dietary CoQ10 exerts a strong antioxidant effect and prevents development of ascites syndrome. Significant reduction in incidence of ascites (Faraji et al. 2019; Sharifi et al. 2016) and ascites induced mortality have been reported by CoQ10 supplementation in diet of broiler chicken (Geng et al., 2004).

On similar lines the herbal supplements containing naturally occurring phytochemicals exhibit a potential for modulating metabolic disorders such as ascites syndrome. The rich pool of bioactive compounds in wild celery (*Kelussia odoratissima*) have been shown to exert strong suppression on pulmonary hypertension and ascites in broiler chicken (Ahmadipour et al. 2015). The ethanol extract of elecampane (*Inula helenium* L.) rhizome, rich in sesquiterpenes, and root extract of *Prosopis farcta*, rich in flavonoids, have prevented ascites in broiler chicken (Abolfathi et al. 2021). The supplementation of purslane (*Portulaca oleracea* L.) powder, rich in phenolic alkaloids and alpha-linolenic acid, has reduced the incidence of ascites in broiler chicken with improved antioxidant status (Habibian et al. 2017; Shirzadi et al. 2020).

5. Conclusions

The propriety dietary formulation, Ascitox Q_{10} , has significantly reduced the mortality of broiler chicken and thus can be valued as a fruitful nutritional intervention in controlling ascites induced mortality in broiler chicken with improved growth performance at commercial farms.

Declarations

Funding: None

Conflict of interest: The author declares no conflict of interest **Ethics approval**: Not applicable

Acknowledgements: The authors wish to express gratitude to IMN Pyrophos which provided Ascitox $-Q_{10}$, the associated field vets, and the farmers

References

- Abolfathi ME, Tabeidian SA, Shahraki AD, Tabatabaei SN, Habibian M. (2021). Ethanol extract of elecampane (*Inula helenium* L.) rhizome attenuates experimental cold-induced ascites (pulmonary hypertension syndrome) in broiler chickens. Animal Feed Science and Technology 272: 114755.
- Acar N, Sizemore FG, Leach GR, Wideman JRF, Owen RL, Barbato GF. (1995). Growth of broiler chickens in response to feed restriction regimens to reduce ascites. Poultry Science 74: 833-843.
- Ahmadipour B, Hassanpour H, Asadi E, Khajali F, Rafiei F, Khajali F. (2015). *Kelussia odoratissima* Mozzaf – A promising medicinal herb to prevent pulmonary hypertension in broiler chickens reared at high altitude. Journal of Ethnopharmacology 159: 49– 54.
- Ahmed M, Biswas A, Roy BG, Srivastava RB. (2013). Most frequently encountered problems during hatching at cold arid high altitude region like Ladakh in India: Causes and its remedies. World's Poultry Science Journal 69: 897-902.
- Al-Zahrani K, Licknack T, Watson DL, Anthony NB, Rhoads DD. (2019). Further investigation of mitochondrial biogenesis and gene expression of key regulators in ascites- susceptible and ascites- resistant broiler research lines. PLoS One 14(3): e0205480.
- Aviagen (2009). Effective Management Practices to reduce the incidence of ascites in broilers. Arbor acres service bulletin. Available online at <u>http://ap.aviagen.com/assets/Tech_Center/</u><u>AA_Technical_Articles/AAServiceBulletinAscites.pdf</u>
- Balog JM. (2003). Ascites syndrome (pulmonary hypertension syndrome) in broiler chickens: Are we seeing the light at the end of the tunnel? Avian and Poultry Biology Review 14(3): 99–126.
- Biswas A, Bharti VK, Raj T, Kumar A, Srivastava RB. (2012). Effects of dietary vitamin E and selenium on growth performance of growing broiler chicken reared at high altitude. Indian Journal of Poultry Science 47: 118-120.
- Biswas A. (2019). Pulmonary hypertension syndrome in broiler chickens: a review. Veterinarski Arhiv 89(5): 723-734.
- Bottje WB, Wideman JRRF. (1995). Potential role of free radicals in the pathogenesis of pulmonary hypertension syndrome. Poultry and Avian Biology Reviews 6(3): 221–231.
- Dahal GK (2011). Ascites (water belly) in broiler chickens during winter season. engormix. <u>https://en.engormix.com/poultry-industry/articles/ascites-in-broiler-chickens-t35214.htm</u>
- Decuypere E, Buys N, Buyse J. (2000). Ascites in broiler chickens: exogenous and endogenous structural and functional causal factors. Worlds Poultry Science Journal 56: 367-377.
- Decuypere E, Hassanzadeh M, Buys N. (2005). Further insights into the susceptibility of broilers to ascites. The Veterinary Journal 169: 319-320.
- Druyan S. (2012). Ascites syndrome in broiler chickens A physiological syndrome affected by Red blood cell. In: Moschandreou TE, editor, Blood cell - An overview of studies in hematology. IntechOpen, London. Available from: <u>https://www.intechopen.com/chapters/39117</u>

- Faraji, M, Karimi Dehkordi S, Zamiani Moghadam AK, Ahmadipour B, Khajali F. (2019). Combined effects of guanidino acetic acid, coenzyme Q₁₀ and taurine on growth performance, gene expression and ascites mortality in broiler chickens. Journal of Animal Physiology and Animal Nutrition 103: 162–169.
- Fedde MR, Weigle GE, Wideman RF. (1998). Influence of feed deprivation on ventilation and gas exchange in broilers: relationship to pulmonary hypertension syndrome. Poultry Science 77: 1704-1710.
- Geng AL, Guo YM, Yang Y. (2004). Reduction of ascites mortality in broilers by Coenzyme Q₁₀. Poultry Science 83: 1587-1593.
- Grygorczyk R, Orlov SN. (2017). Effects of hypoxia on erythrocyte membrane properties-Implications for intravascular hemolysis and purinergic control of blood flow. Frontiers in Physiology 8: 1110. https://doi.org/10.3389/fphys.2017.01110
- Habibian M, Sadeghi G, Karimi A. (2017). Effects of purslane (*Portulaca oleracea* L.) powder on growth performance, blood indices, and antioxidant status in broiler chickens with triiodothyronine-induced ascites. Archives of Animal Breeding 60: 315–325.
- Halliwell B, Gutteridge JMC. (1990). Role of free radicals and catalytic metal ions in human disease: An overview. Methods in Enzymology 186: 185–204.
- Hassanzadeh M, Buys N, Dewil E, Rahimi G, Decuypere E. (1997). The prophylactic effect of vitamin C supplementation on broiler ascites incidence and plasma thyroid hormone concentration. Avian Pathology 26: 33–44.
- Havenstein GB, Ferket PR, Scheideler SE, Larson BT. (1994). Growth, livability, and feed conversion of 1957 vs 1991 broilers when fed typical 1957 and 1991 broiler diets. Poultry Science 73: 1785-1794.
- Julian RJ. (1990). Cardiovascular disease. In: Jordan FTW, editor, Poultry diseases (3rd ed). Bailliere Tindall, London, United Kingdom, pp. 345–353.
- Julian RJ. (1993). Ascites in poultry. Avian Pathology 22: 419-454.
- Julian RJ. (2000). Physiological management and environmental triggers of the ascites syndrome: A review. Avian Pathology 29: 519-527.
- Julian RJ. (2005). Production and growth related disorders and other metabolic diseases of poultry - A review. The Veterinary Journal 169: 350-369
- Kalmar ID, Vanrompay D, Janssens GPJ (2013). Broiler ascites syndrome: Collateral damage from efficient feed to meat conversion. The Veterinary Journal 197(2): 169-174.
- Khajali F, Fahimi S. (2010). Influence of dietary fat source and supplementary α -tocopheryl acetate on pulmonary hypertension and lipid peroxidation in broilers. Journal of Animal Physiology and Animal Nutrition 94: 767–772.
- Kornfeld OS, Hwang S, Disatnik MH, Chen CH, Qvit N, Mochly-Rosen D. (2015). Mitochondrial reactive oxygen species at the heart of the matter: New therapeutic approaches for cardiovascular diseases. Circulation Research 116(11): 1783-1799.
- Lorenzoni AG, Ruiz-Feria CA. (2006). Effects of vitamin E and Arginine on cardiopulmonary function and acites parameters in broiler chickens reared under subnormal temperatures. Poultry Science 85: 2241-2250.

- Luger D, Shinder D, Wolfenson D, Yahav S. (2003). Erythropoiesis regulation during the development of ascites syndrome in broiler chickens: A possible role of corticosterone on egg production. Journal of Animal Science 81: 784-790.
- Mirsalimi SM, Julian RJ. (1991). Reduced erythrocyte deformability as a possible contributing factor to pulmonary hypertension and ascites in broiler chickens. Avian Diseases 35: 374-379.
- Moghaddam AZ, Hamzekolaei MM, Khajali F, Hassanpour H. (2017). Role of selenium from different sources in prevention of pulmonary arterial hypertension syndrome in broiler chickens. Biological Trace Elements Research 180: 164-170.
- Mohammadalipour R, Rahmani HR, Jahanian R, Riasi A, Mohammadalipour M, Nili N. (2017). Effect of early feed restriction on physiological responses, performance and ascites incidence in broiler chickens raised in normal or cold environment. Animal 11: 219–226.
- Olkowski AA. (2007). Pathophysiology of heart failure in broiler chickens: Structural, biochemical, and molecular characteristics. Poultry Science 86(5): 999-1005.
- Owen RL, Wideman RF, Leach RM, Cowen BS, Dunn PA, Ford BC. (1994). Effect of age of exposure and dietary acidification or alkalinization on broiler pulmonary hypertension syndrome. Journal of Applied Poultry Research 3(3): 244-252.
- Ozkan SEZE, Malayoglu HB, Yalcin S, Karadaş F, Koçturk S, Cabuk M, Oktay G, Ozdemir S, Ozdemir E, Ergul M. (2007). Dietary vitamin E (α-tocopherol acetate) and selenium supplementation from different sources: Performance, ascites-related variables and antioxidant status in broilers reared at low and optimum temperatures. *British Poultry Science*, 48, 580–593.
- Peoples JN, Saraf A, Ghazal N, Pham TT, Kwong JQ. (2019). Mitochondrial dysfunction and oxidative stress in heart disease. Experimental and Molecular Medicine 51(12): 162.
- Reeves, J. T., G. Ballam, S. Hofmeister, C. Pickett, K. Morris, and A. Peacock. 1991. Improved arterial oxygenation with feed restriction in rapidly growing broiler chickens. Comp. Biochem. Physiol. A. Comp. Physiol. 99: 481-485.
- Roch G, Boulianne M, De Roth L. (2000). Effect of vitamin E and selenium on incidence of ascites, growth performance and blood parameters in cold-stressed broilers. Poultry Science 79(1):41
- Ruiz-Feria CA. (2009). Concurrent supplementation of arginine, vitamin E, and vitamin C improve cardiopulmonary performance in broilers chickens. Poultry Science 88: 526–535.
- Saki AA, Hemati M. (2011). Does nutrition help to alleviate sudden death syndrome in broiler chicken? Global Veterinarian 6(3): 262-268.
- Sharifi MR, Khajali F, Hassanpour H. (2016). Antioxidant supplementation of low-protein diets reduced susceptibility to pulmonary hypertension in broiler chickens raised at high altitude. Journal of Animal Physiology and Animal Nutrition 100: 69–76.
- Shirzadi H, Shariatmadari F, Karimi Torshizi MA, Rahimi S, Masoudi AA, Zaboli GR, Hedayat-Evrigh N. (2020). Plant extract supplementation as a strategy for substituting dietary antibiotics in broiler chickens exposed to low ambient temperature. Archives of Animal Nutrition 74: 206–221.
- Villar-Patino G, Diaz-Cruz A, Avila-Gonzalez E, Guinzberg R, Pablos JL, Pina E. (2002). Effects of dietary supplementation with

vitamin C or vitamin E on cardiac lipid peroxidation and growth performance in broilers at risk of developing ascites syndrome. American Journal of Veterinary Research 63: 673–676.

- Wideman RF, Ismail JRM, Kirby YK, Bottje WG, Varderman RC. (1995). Furosemide reduces the incidence of pulmonary hypertension syndrome (ascites) in broilers exposed to cool environmental temperatures. Poultry Science 74: 314–322.
- Wideman RF, Rhoads DD, Efr GF, Anthony NB. (2013). Pulmonary arterial hypertension (ascites syndrome) in broilers: A review. Poultry Science 92: 64-83.
- Wuyi L. (2016). A trial diagnosis of ascites syndrome in broiler chickens – Case report. Pakistan Journal of Biological Sciences 19: 352-359.
- Xiang RP, Sun WD, Wang JY, Wang XL. (2002). Effect of vitamin C on pulmonary hypertension and muscularization of pulmonary arterioles in broiler. British Poultry Science 43: 705-712.

Citation

Ali I, Naqvi S. (2022). Control of winter ascites in commercial broiler chicken through propriety formulation. Letters in Animal Biology 02(2): 22 – 28.